AUTONOMOUS CHOLESTEROL AND FATTY ACID SYNTHESIS IN HEPATOMAS: DELETION OF THE ADENOSINE 3', 5'-CYCLIC MONOPHOSPHATE CONTROL MECHANISM OF NORMAL LIVER

Lee A. Bricker and Gerald S. Levey

Divisions of General Medicine and Endocrinology and Metabolism Department of Medicine, University of Miami School of Medicine Miami, Florida 33152

Received June 12,1972

Summary: Rat hepatoma slices incubated with cAMP $(5x10^{-3}\text{M})$ or dibutyryl cAMP $(3x10^{-4}\text{M})$, and acetate-2- ^{14}C , demonstrated unsuppressible cholesterol and fatty acid synthesis, compared to controls. This observation contrasts sharply with that of the marked suppression in lipogenesis brought about by cyclic nucleotides in normal liver and indicates a deletion in hepatomas of this regulatory mechanism in lipogenesis.

We have previously reported that cholesterol and fatty acid synthesis in normal liver is suppressible by adenosine 3', 5'-cyclic monophosphate (cAMP) (1) and have suggested the possibility that this is a physiologically meaningful mechanism in the regulation of hepatic lipogenesis. The sequence of events in liver appears to be quite specific, apparently involving an early step in the utilization of acetyl CoA. Similar considerations involving hepatomas presented themselves, since many of these malignant tumors likewise are highly active producers of sterols (2). Specific defects in regulation of tumor sterologenesis have been well described previously (3,4), involving, primarily, the conversion of β -hydroxy- β -methylglutaryl CoA to mevalonate. The purpose of this investigation was to examine the possibility that a separate regulatory defect in lipogenesis related to cAMP exists in these neoplasms.

MATERIALS AND METHODS

The experimental protocol has been described previously (1). The procedure utilized tissue slices incubated in Krebs' bicarbonate buffer with 2 μ Ci acetate-2-14C, and either a final concentration of cAMP of $5 \times 10^{-3} \text{M}$, or of dibutyryl cAMP, $3 \times 10^{-4} \text{M}$. Controls were identical but with no added nucleotide. Slices of liver and hepatoma were 0.33 mm thick; 350 mg of slices were used in the incubations,

carried out at 37° C for two hours in a metabolic shaker at 100 oscillations per minute.

All experiments involved male rats of the ACI/f Mai strain, maintained on Purina rat chow. Two strains of transplantable hepatoma were studied: 1) Morris hepatoma #3924A, a poorly-differentiated, rapidly-growing adenocarcinoma of liver, and 2) Morris hepatoma #9121, a so-called "minimal-deviation" hepatoma, which grows slowly and is very active in sterol synthesis. Both hepatoma types were implanted in the right thighs of the host animals.

Digitonin-precipitable sterols, fatty acids, and metabolically-generated CO2 were assayed as described previously (1). Radioactivity was determined with a Packard liquid scintillation spectrometer, model 3390, using a scintillation fluid constructed as outlined elsewhere (5).

RESULTS AND DISCUSSION

Table 1 compares the effects of cAMP and dibutyryl cAMP on incorporation of acetate-2-14C into cholesterol (as digitonin-precipitable sterol), fatty acid, and 14CO2 by Morris hepatoma #3924A, the liver of the tumor-bearing animal, and the liver of a comparable, tumor-free animal. The tumor, in contrast to the normal liver from the tumor-free animal, showed no suppression in sterologenesis or fatty acid synthesis with either cyclic nucleotide. Both normal liver and, to a lesser extent, the liver from the tumor-bearing animal, demonstrated inhibition in lipogenesis resulting from incubation with both cAMP and dibutyryl cAMP. In no instance was 14CO2 evolution depressed. Table 2 shows the results of a similar experiment on Morris hepatoma #9121. The results are qualitatively similar. The capacity for regulation of lipogenesis by cAMP is absent in both the undifferentiated (#3924A) and the "minimal deviation" (#9121) hepatomas.

The observed failure of cAMP to suppress lipogenesis in hepatomas contrasts with our previously reported findings in normal liver (1). Taken together, these data correlate well with those of Granner (6), demonstrating the absence of cAMP binding protein in hepatoma cells and its presence in normal liver cells. Presumably absence of this binding protein would prevent the normal response to cAMP.

TABLE 1

		DPS*	FA	14 _{C02}	
Normal liver-(tumor-free animal)					
	Control	21717±2421	7596±1408	23956±1854	
	Dibutyryl cAMP	5248±803	690±112		
	cAMP	5578±839	936±244	29938±15981	
Morris hepatoma #3924A.					
	Control	27805±5343	1985±354	15885±1210	
	Dibutyry1 cAMP	27762±11555	1699±1760	17252±5504	
	cAMP	23461±4949	1739±499	17171±7128	
Liver-(tumor-bearing animal)					
	Control	3834±881	921±197	22184±4337	
	Dibutyryl cAMP	2117±450	542±136	18640±1275	
	cAMP	2569±457	468±79	34997±1050	

Table 1: Effects of cAMP and dibutyryl cAMP on incorporation of ^{14}C -acetate into digitonin-precipitable sterol, fatty acid and $^{14}\text{CO}_2$ by tissue slices of liver and Morris hepatoma #3924A. Data represents cpm incorporated by 350 mg slices/2 hr., as mean \pm S.E. of triplicate analyses. Relative incorporation of acetate-2-14C into these three fractions may be obtained by multiplying cpm value for fatty acids by 4, and for CO₂ by 10. Final concentrations of cAMP and dibutyryl cAMP are as indicated in the text.

ACKNOWLEDGEMENTS

Dr. Levey is an investigator of the Howard Hughes Medical Institute.

We are indebted to Mrs. Patricia L. Kozlovskis and Mrs. Jacquelyn Glasser for able technical assistance, and to Mrs. Brigita Rowe for preparation of the

^{*} Digitonin-precipitable sterol (predominantly cholesterol)

TABLE 2

	DPS*	<u>FA</u>	14 _{C02}		
Normal liver-(tumor-free animal)					
Control	12190±2462	8342±715	24072±7780		
cAMP	5745±766	1333±182	20641±3996		
Morris hepatoma #9121					
Control	17240±462	1827±472	7251±1105		
CAMP	22376±2585	1496±122	7518±672		
Liver-(tumor-bearing animal)					
Control	6034±667	2216±770	41913±5223		
cAMP	3895±906	831±268	37655±8969		

Table 2: Effects of cAMP on incorporation of ^{14}C -acetate into digitonin-precipitable sterol, fatty acid, and $^{14}\text{CO}_2$ by tissue slices of liver and Morris hepatoma #9121. Data represents cpm incorporated by 350 mg. of slices/2 hr., as mean \pm S.E. of triplicate analyses. Relative incorporations of acetate-2- ^{14}C into these three fractions may be obtained as in the legend of Table 1.

manuscript. This work was supported by U.S. Public Health Grants CA 11969-02, HE 13715-02, HL 14141-02, Florida Heart Association Grant 72 AG 20, and an award from the Damon Runyon Cancer Fund (DRG 1120).

REFERENCES

- 1. L.A. Bricker and G.S. Levey, J. Biol. Chem., in press
- 2. M.D. Siperstein and V.M. Fagan, Cancer Res. 24, 1108 (1964)
- 3. M.D. Siperstein, V.M. Fagan, H.P. Morris, Cancer Res. 26, 7 (1966)
- 4. L.A. Bricker, H.P. Morris, M.D. Siperstein, J. Clin. Invest. 51, 206 (1972)
- 5. L.A. Bricker, H.J. Weis, M.D. Siperstein, J. Clin. Invest. 51, 197 (1972)
- 6. D.K. Granner, Biochem. Biophys. Res. Commun. 46, 1516 (1972)

^{*} Digitonin-precipitable sterol (predominantly cholesterol)